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Production, Respiration, and Overall Carbon Balance in an Old-growth Pseudotsuga-Tsuga Forest Ecosystem

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ABSTRACT

Ground-based measurements of stores, growth, mortality, litterfall, respiration, and decomposition were conducted in an old-growth forest at Wind River Experimental Forest, Washington, USA. These measurements were used to estimate gross primary production (GPP) and net primary production (NPP); autotrophic respiration (Ra) and heterotrophic (Rh) respiration; and net ecosystem production (NEP). Monte Carlo methods were used to calculate uncertainty (expressed as ± 2 standard deviations of 200–400 calculations). Live carbon (C) stores were 39,800 g C m⁻² (34,800–44,800 g C m⁻²). The store of C in detritus and mineral soil was 22,092 g C m⁻² (20,600–23,600 g C m⁻²), and the total C stores were 61,899 g C m⁻² (56,600–67,700 g C m⁻²). Total NPP was 597 g C m⁻² y⁻¹ (453 to 741 g C m⁻² y⁻¹). Ra was 1309 g C m⁻² y⁻¹ (845–1773 g C m⁻² y⁻¹), indicating a GPP of 1906 g C m⁻² y⁻¹ (1444–2368 g C m⁻² y⁻¹). Rh, including the respiration of heart rots in tree boles, was 577 g C m⁻² y⁻¹ (479–675 g C m⁻² y⁻¹). Long-term NEP was estimated to be +20 g C m⁻² y⁻¹ (-116 to +156 g C m⁻² y⁻¹), indicating this stand might be a small sink. These estimates contrast with the larger sink estimated at the same site using eddy-flux methods. Several hypotheses to explain this discrepancy were explored, including (a) undetected biomass increases, (b) underestimates of NPP, (c) unmeasured losses, and (d) a temporal mismatch between the two sets of measurements. The last hypothesis appears the most likely.

Key words: autotrophic respiration; carbon flux; carbon stores; decomposition; gross primary production (GPP); heterotrophic respiration; net ecosystem production (NEP); net primary production (NPP).

INTRODUCTION

Net ecosystem production (NEP) of terrestrial ecosystems is a key process to understand when actively managing the carbon (C) cycle. Despite the need to understand how and why this rate varies in space and time, most ecosystem studies have focused on net primary productivity (NPP). While knowledge of controls of NPP is important, several other processes including heterotrophic respiration (Rh) and disturbance also have to be understood before NEP can be determined (Randerson and others 2002).
There are several approaches to estimating NEP in forests. Long-term observations of growth, mortality, litterfall, harvest, and decomposition, coupled with detritus and soil stores, as well as short-term measurements of respiration can be used in the "aggregated" flux method (Grier and Logan 1977; Law and others 2000). Alternatively, NEP can be estimated by measuring the change in total C stores over time by using the delta-stores method (Turner and others 1999). Simulation models have also been used to predict changes in NEP, either in response to climate change, carbon dioxide (CO₂) increases, and increased nitrogen deposition [for example, see Lloyd (1999)] or disturbance, including timber harvest [for example, see Harmon and others (1990)]. These models are largely based on a synthesis of ground-based measurements similar to the aggregated-flux method. Most recently, estimates of NEP for entire ecosystems (that is, not one part at a time) have been derived from micrometerologic methods such as eddy covariance (Wofsey and others 1993; Goulden and others 1996). Given that each of these methods has developed at different times and places, there have been very few comparisons of all methods at one place and time. Research currently being conducted at the Wind River Experimental Forest, Washington, USA, provides such an opportunity.

Our objectives in this report are (a) to use the ground-based, aggregated-flux method to estimate gross primary production (GPP) and net primary production (NPP), respiration of autotrophs (Rₐ) and heterotrophs (Rₕ) of an old-growth forest at Wind River; (b) to use these numbers to estimate the net ecosystem production (NEP) of this forest; and (c) to compare our long-term, ground-based estimates of NEP with those derived by eddy covariance (Paw U and others 2004) and simulation models [that is, the Soil–Plant–Atmosphere model (Winner and others 2004)]. We then evaluate alternative hypotheses that might resolve the differences (if any) in the estimates derived from these three independent methods.

**Study Area**

This study was conducted at the Wind River Canopy Crane Research Facility (WRCCRF). The site is typical of old-growth forests west of the Cascades (Shaw and others 2004). A detailed description of this old-growth stand is presented by Parker and colleagues (2004). The forest is classified as a western hemlock–salal cover type and is estimated to be approximately 500 years old. Dominant tree species are Douglas-fir (*Pseudotsuga menziesii*) and western hemlock (*Tsuga heterophylla*). Dominant understory shrub species are vine maple (*Acer circinatum*), salal (*Gaultheria shallon*), and dwarf Oregon grape (*Berberis nervosa*). The climate is characteristic of a temperate winter-wet, summer-dry climate. Annual precipitation totals 2467 mm, with less than 10% occurring between June and September. Mean annual temperature is 8.7°C. Soils are of the Stabler series, coarse textured and developed on 2–3 m of volcanic ejecta over basalt bedrock. Texture ranges from shotty loam to clay with coarse particles in the top 1 m averaging 3% of the soil volume.

**Methods**

**Live Biomass**

**Tree Biomass.** Tree biomass, mortality, and ingrowth were estimated around the Wind River canopy crane in a 4-ha plot (hereafter, the crane plot) that was established in 1994 and divided into four 1-ha quadrants, each of which were further divided into 16 numbered 25-m × 25-m subplots. All trees larger than 5-cm DBH (diameter at breast height) were measured for diameter and height as well as tagged with aluminum tags at breast height (1.3 m). Diameter was measured with a tape to the nearest 0.1 cm. Annual surveys were conducted between 1995 and 1999 to determine the trees that died. All surviving trees and those that grew into the minimum diameter class (that is, ingrowth) were measured in summer 1999. Biomass of all live tree parts and volume for thebole were calculated using allometric equations (Gholz and others 1979; Means and others 1994). Species-specific allometric equations were used when available, and substitutions for some minor species (for example, *Abies grandifolia* and *Taxus brevifolia*) were used. Coarse-root allometric equations were used for roots larger than 5 mm in diameter. The mass of roots 2–5 mm in diameter from fine-root cores (see below) was added to the allometric equation estimates to calculate the total mass of coarse roots. Leaf mass was estimated using a sapwood area-based estimate using DBH-sapwood thickness and leaf-area relationships developed for the H. J. Andrews Experimental Forest in the central Cascades of Oregon (Gholz and others 1976; Waring and others 1982; Means and others 1999). Sapwood volume was estimated from equations developed by Harcombe and colleagues (1990) that predict the proportion of the total bole in sapwood from DBH.
Table 1. Estimated Stores of Carbon and Rates of Production Associated with Live Biomass; Mean (Standard Error)

<table>
<thead>
<tr>
<th>Pool</th>
<th>Store</th>
<th>Δ Stores</th>
<th>Mortality/Litterfall</th>
<th>NPP</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>g C m⁻²</td>
<td>g C m⁻² y⁻¹</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Stem sapwood</td>
<td>6567 (198)</td>
<td>2 (1)</td>
<td>30 (9)</td>
<td>32 (9)</td>
</tr>
<tr>
<td>Stem heartwood</td>
<td>15,351 (1151)</td>
<td>26 (2)</td>
<td>50 (13)</td>
<td>76 (13)²</td>
</tr>
<tr>
<td>Stem bark</td>
<td>3337 (263)</td>
<td>2 (4)</td>
<td>12 (2)</td>
<td>14 (3)</td>
</tr>
<tr>
<td>Live branches</td>
<td>4489 (112)</td>
<td>8 (10)</td>
<td>100 (34)</td>
<td>108 (34)</td>
</tr>
<tr>
<td>Dead branches</td>
<td>318 (20)</td>
<td>0 (0)</td>
<td>3 (1)</td>
<td>3 (1)</td>
</tr>
<tr>
<td>Tree foliage</td>
<td>941 (322)</td>
<td>0 (0)</td>
<td>135 (12)</td>
<td>150 (14)²</td>
</tr>
<tr>
<td>Coarse roots</td>
<td>8122 (639)</td>
<td>21 (7)</td>
<td>30 (5)</td>
<td>51 (7)</td>
</tr>
<tr>
<td>Fine roots</td>
<td>362 (26)</td>
<td>0 (0)</td>
<td>91 (16)²</td>
<td>91 (16)</td>
</tr>
<tr>
<td>Understory shrubs</td>
<td>144 (37)</td>
<td>0 (0)</td>
<td>26 (5)²</td>
<td>26 (5)</td>
</tr>
<tr>
<td>Understory herbs</td>
<td>76 (8)</td>
<td>0 (0)</td>
<td>40 (8)²</td>
<td>40 (8)</td>
</tr>
<tr>
<td>Epiphytes</td>
<td>100 (25)</td>
<td>0 (0)</td>
<td>6 (1)</td>
<td>6 (1)</td>
</tr>
<tr>
<td>Total</td>
<td>39,807 (2479)</td>
<td>59 (24)</td>
<td>523 (69)</td>
<td>597 (72)²</td>
</tr>
</tbody>
</table>

*The net change in stocks in these pools was assumed to be zero.
*Assumed that all leaves were from shrubs that died each year and that 0.5%–1.0% of the stems died. Litter traps indicate that the value of shrub litterfall may be as low as 1 g C m⁻² y⁻¹.
*Assumed that litterfall from herbs was 40%–60% of live stores to account for the fact some small woody-stemmed, evergreen plants are included in the herb category.
*Assumes no heart rot is present.
*Includes grazing of 15 g C m⁻² y⁻¹.
*Assumes 20%–30% of fine roots die annually.
C. carbon; and NPP, net primary production.

**Fine Roots.** In October 2000, 20 soil cores of 5-cm diameter to a depth of 1 m were removed to estimate biomass of fine roots less than 2 mm in diameter. In each 1-ha quadrant of the crane plot, five cores were sampled at random distances along transects placed diagonally across the quadrant. Organic horizons were sorted by hand to remove live and dead fine roots. Mineral soil was subdivided into 20-cm depths and then washed using a root elutor to separate roots. Roots were sorted into size classes and live versus dead, oven dried at 55°C, and weighed. Subsamples of root material were placed in an oven at 550°C for 4 h to determine ash-free dry weights. Means and standard errors were calculated using all 20 samples as a basis.

**Understory Plants.** The aboveground biomass of understory shrubs and trees larger than 5-cm DBH was estimated at 21 locations within the T. T. Munger Research Natural Area (RNA) by recording their diameter at the base (except salal and Oregon grape, which were treated as herbs) within a 25 x 1-m belt transect at each location. This RNA surrounds the crane plot (Shaw and others 2004). These sampling locations were in existing plots systematically placed throughout the RNA at 100-m spacing (Smithwick and others 2002). The cover of mosses, herbs, salal, and Oregon grape was determined in 25-, 20-, by 50-cm microplots systematically placed along each of the 25-m belt transects. The biomass of understory plants was calculated using allometric equations (Means and others 1994). In cases where equations for a species (particularly herbaceous ones) did not exist, equations from similar species were used.

**Epiphytes.** The biomass of epiphytes was based on work by McCune (1993) and McCune and colleagues (1997) from aerial and ground-based surveys conducted at the Wind River site and other old-growth Douglas-fir forests in the Pacific Northwest region. The most precise estimates of epiphyte biomass were for lichens. We therefore multiplied the estimated biomass of lichens by 2, given bryophytes and lichens are of approximately equal abundance (McCune 1993).

**Net Primary Production**

NPP estimates of the woody parts of trees (that is, sapwood, heartwood, bark, branches, and coarse roots) were assumed equivalent to the change in live stores, the losses from mortality (that is, entire death of trees), and ingrowth gains from trees that grew into the minimum size class:

\[ NPP_w = \Delta \text{stores}_w + \text{mortality}_w + \text{ingrowth}_w \]

Where \( w \) represents a woody tree part (Clark and others 2001). Note that \( \Delta \text{stores} \) for heartwood did not account for the possible loss from heart-rot decomposition. We accounted for those losses under heterotrophic respiration (see below). For nonwoody, aboveground components, NPP was
Table 2. Estimated Rates of Production and Respiration Associated with Live Biomass; Mean (Standard Error)

<table>
<thead>
<tr>
<th>Pool</th>
<th>$R_a$</th>
<th>NPP</th>
<th>GPP</th>
</tr>
</thead>
<tbody>
<tr>
<td>Stem sapwood</td>
<td>153 (11)</td>
<td>32 (9)</td>
<td>185 (13)</td>
</tr>
<tr>
<td>Stem heartwood</td>
<td>NA</td>
<td>76 (13)$^a$</td>
<td>76 (11)</td>
</tr>
<tr>
<td>Stem bark</td>
<td>NA$^b$</td>
<td>14 (3)</td>
<td>14 (3)</td>
</tr>
<tr>
<td>Live branches</td>
<td>7 (0.5)</td>
<td>108 (34)</td>
<td>115 (32)</td>
</tr>
<tr>
<td>Dead branches</td>
<td>NA$^c$</td>
<td>3 (1)</td>
<td>3 (1)</td>
</tr>
<tr>
<td>Tree foliage</td>
<td>577 (233)</td>
<td>150 (14)$^d$</td>
<td>727 (218)</td>
</tr>
<tr>
<td>Coarse roots</td>
<td>167 (21)</td>
<td>51 (7)</td>
<td>218 (20)</td>
</tr>
<tr>
<td>Fine roots</td>
<td>274 (30)</td>
<td>91 (16)</td>
<td>365 (32)</td>
</tr>
<tr>
<td>Understory shrubs</td>
<td>10 (4)</td>
<td>26 (5)</td>
<td>36 (6)</td>
</tr>
<tr>
<td>Understory herbs</td>
<td>57 (15)</td>
<td>40 (8)</td>
<td>97 (16)</td>
</tr>
<tr>
<td>Epiphytes</td>
<td>64 (19)</td>
<td>6 (1)</td>
<td>70 (17)</td>
</tr>
<tr>
<td>Total</td>
<td>1309 (232)</td>
<td>597 (72)</td>
<td>1906 (231)</td>
</tr>
</tbody>
</table>

$^a$Does not account for possible losses from heart rot in range of 0–46 g C m$^{-2}$ y$^{-1}$.
$^b$Inner bark accounted for in sapwood calculation.
$^c$Estimated for understorey in Table 3.
$^d$Includes aboveground grazing of 15 g C m$^{-2}$ y$^{-1}$; the litterfall portion was 135 (12) g C m$^{-2}$ y$^{-1}$, mean (standard error).

Estimated from litterfall and tree foliage grazing estimates. A total of 20 litter traps, 40 × 40-cm squares lined with fine mesh, were used to collect fine litterfall between June 1997 and December 1999. The 0.5-m-tall traps were placed randomly at five fixed locations in each of the 1-ha quadrants of the 4-ha crane plot. Samples were collected monthly (except when snow depth exceeded trap depth) and then oven-dried and weighed to the nearest 0.001 g. and sorted into nine categories: green conifer needles; brown conifer needles; nonconifer leaves; twigs (woody); cones, seeds, other reproductive parts; cyanolichens; other lichens; moss; and miscellaneous materials. Woody material larger than 1 cm in diameter was excluded, and accounted for under mortality. To estimate spatial variance, the mean litterfall from each of the 20 traps was used for the 2.5-year period of observation. For understory herbs, we estimated aboveground NPP assuming A stores was 0 and that 40%–60% of the aboveground biomass died annually. A portion less than unity was used because approximately 50% of the herb layer was comprised of woody evergreen shrubs with a leaf life span of 2 years. For shrubs, we assumed the litterfall portion of NPP was captured in litter traps and was represented by nonconifer leaves. The mortality of woody shrubs was not measured, but was estimated by assuming 1% of the stems died annually. Fine-root production was estimated by assuming fine-root biomass was not increasing and that annual fine-root mortality was 20%–30%.

This preliminary mortality estimate was from 40 minirhizotron tubes in the crane plot that were visited monthly over 1999 and based on standard methods (Fahey and others 1999).

We estimated aboveground grazing of trees from visual surveys of foliage damage in the canopy crane plot conducted in 1995, 1996, and 1999. We did not account for belowground herbivory (other than that associated with fine-root mortality) and assumed the belowground herbivory flux to be zero. In 1995 and 1996, the fraction of the current-year foliage that was damaged by herbivory and other unknown causes was noted in a systematic survey of the lower, middle, and upper canopy by using the canopy crane for access. We assumed that all this damage was from herbivory. In 1999, herbivory damage to all age classes of foliage was visually estimated using a similar survey method as earlier. To estimate the mass of herbivory, we multiplied the fraction damaged by the appropriate foliage mass (that is, current-year mass for 1995 and 1996 and total mass for 1999). We assumed that all herbivory resulted in respiration, either directly or in the death and decomposition of herbivores. Because the majority of aboveground herbivory is probably from insects, this assumption is quite reasonable.

Autotrophic Respiration

Autotrophic respiration was estimated as a rate constant times the biomass times a temperature
Table 3. Estimated Stores of Carbon, Rate Constants, and Fluxes Associated with Grazers, Heart Rot, Detritus and Soil Pools; Mean (Standard Error)

<table>
<thead>
<tr>
<th>Pool</th>
<th>Store</th>
<th>Rate Constant</th>
<th>(R_h)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Logs Pseudotsuga</td>
<td>1920 (170)</td>
<td>0.014 (0.006)</td>
<td>27 (11)</td>
</tr>
<tr>
<td>Thuja</td>
<td>50 (20)</td>
<td>0.007 (0.003)</td>
<td>0.3 (0.2)</td>
</tr>
<tr>
<td>Other</td>
<td>2220 (210)</td>
<td>0.018 (0.003)</td>
<td>40 (7)</td>
</tr>
<tr>
<td>Subtotal</td>
<td>4190 (310)</td>
<td>0.016</td>
<td>67 (12)</td>
</tr>
<tr>
<td>Snags Pseudotsuga</td>
<td>2170 (390)</td>
<td>0.021 (0.009)</td>
<td>46 (19)</td>
</tr>
<tr>
<td>Thuja</td>
<td>10 (1)</td>
<td>0.011 (0.005)</td>
<td>0.1 (0.1)</td>
</tr>
<tr>
<td>Other</td>
<td>710 (140)</td>
<td>0.028 (0.004)</td>
<td>20 (4)</td>
</tr>
<tr>
<td>Subtotal</td>
<td>2890 (430)</td>
<td>0.023</td>
<td>66 (19)</td>
</tr>
<tr>
<td>Fine woody debris Downed</td>
<td>450 (70)</td>
<td>0.064 (0.010)</td>
<td>29 (5)</td>
</tr>
<tr>
<td>Dead on snags</td>
<td>150 (40)(^a)</td>
<td>0.050 (0.001)</td>
<td>8 (2)</td>
</tr>
<tr>
<td>Dead attached</td>
<td>320 (20)</td>
<td>0.050 (0.001)</td>
<td>16 (1)</td>
</tr>
<tr>
<td>Subtotal</td>
<td>920 (82)</td>
<td>0.058</td>
<td>53 (7)</td>
</tr>
<tr>
<td>Litter</td>
<td>1780 (40)</td>
<td>0.105 (0.020)</td>
<td>187 (37)</td>
</tr>
<tr>
<td>Dead fine roots</td>
<td>476 (112)</td>
<td>0.191 (0.013)</td>
<td>91 (21)</td>
</tr>
<tr>
<td>Decomposed wood</td>
<td>940 (120)</td>
<td>0.007 (0.003)</td>
<td>7 (3)</td>
</tr>
<tr>
<td>Dead coarse roots Pseudotsuga</td>
<td>927 (80)(^b)</td>
<td>0.011 (0.003)</td>
<td>10 (3)</td>
</tr>
<tr>
<td>Thuja</td>
<td>10 (0)(^b)</td>
<td>0.019 (0.002)</td>
<td>0.2 (0.1)</td>
</tr>
<tr>
<td>Other</td>
<td>659 (60)(^b)</td>
<td>0.019 (0.002)</td>
<td>12 (1)</td>
</tr>
<tr>
<td>Subtotal</td>
<td>1596 (103)</td>
<td>0.014</td>
<td>22 (1)</td>
</tr>
<tr>
<td>Mineral soil C to 100 cm</td>
<td>9300 (530)</td>
<td>0.005 (0.0013)</td>
<td>46 (12)</td>
</tr>
<tr>
<td>Grazer respiration</td>
<td></td>
<td>15 (2)(^1)</td>
<td></td>
</tr>
<tr>
<td>Stem heart rot</td>
<td></td>
<td>23 (13)</td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>22,092 (750)</td>
<td>0.026</td>
<td>577 (49)</td>
</tr>
</tbody>
</table>

\(^a\)Estimated from a ratio of dead branches to dead boles. Only class 1 and 2 snags were considered to have dead branches.

\(^b\)Estimated from the ratio of dead roots to sap and log mass.

\(^c\)Based on rate constants for logs adjusted upward to match the mean difference measured by Graham (1982).

\(^d\)Based on rate constants for downed fine woody debris and adjusted according to difference between drowned and suspended branches in time series experiments.

\(^e\)Estimated through extrapolation of long-term data on decomposition.

\(^f\)Based on changes in density of class 3 logs over a 15-year period.

\(^g\)Assumed to be the same as Tsuga.

\(^h\)Assumed to be half the rate of Pseudotsuga.

\(^1\)All grazing consumption assumed to be resired.

\(^2\)A rate constant is the proportion of each pool turning over each year. \(R_h\) heterotrophic respiration.

adjustment. Measurements of foliar respiration, taken at daytime after shading, were the only autotrophic respiration measurements made at the WRCRDF. Therefore, our estimates of \(R_a\) are preliminary and rely on measurements of the same species at other locations. The effect of temperature on diurnal and annual respiration totals was estimated (Agren and Axelsson 1980), with a mean annual temperature of 8.7°C; diurnal temperature amplitudes of 5°C for air and foliage temperature and 1°C for soil temperature; and 15°C for annual temperature amplitude. The measured response of respiration to temperature was used when available, but a Q_{10} of 2 was assumed otherwise (Amthor 1989). We estimated both maintenance (\(R_m\)) and construction (\(R_c\)) respiration (Ryan 1991a). For woody \(R_m\), we used rates per unit sapwood C and assumed that branches, coarse roots, and shrubs were 100% sapwood. Large branches and coarse roots likely have heartwood for these large trees, so this assumption may overestimate sapwood biomass. However, this is offset by the fact that respiration rates for branches and coarse roots are likely to be greater than for stems (Sprugel 1990; Ryan and others 1995, 1996). We used respiration rates measured in the autumn after growth had ceased, and we assumed such rates apply for the entire year. We estimated foliage maintenance respiration for night only, assuming night temperatures averaged 3.7°C (mean annual temperature minus diurnal amplitude) and that yearly foliar respiration rates were the average of rates measured in summer and autumn. We assumed that respiration rates per mass of herbs and epiphytes were the same as conifer foliage. We estimated construction respiration as 25% of NPP for all live components (Ryan 1991b). For all these terms, we assumed that the respiration rates were accurate to
within ±10% of the actual number. This variation, and that caused by variation in the pool sizes, were used to determine a standard error for all \( R_a \) fluxes.

**Gross Primary Production**

GPP was estimated by adding the autotrophic respiration fluxes to the net primary production (NPP) fluxes:

\[
GPP = NPP + R_a
\]

Respiration losses associated with heart rot within living trees was assumed to be part of heterotrophic respiration (see below).

**Detritus and Mineral Soil: Stores and Decomposition**

*Coarse Woody Detritus.* Downed coarse woody detritus (larger than 10 cm in diameter at the large end) was measured using the line-intercept method (Harmon and Sexton 1996). The diameter, species, and decay class of all downed wood crossing the boundaries between all the 25 × 25-m subplots was measured within the 4 ha that was inventoried for live trees and in another 8 ha of plots that had been established next to the crane plot. All the plots are contiguous, forming a 12-ha sample. All standing dead trees larger than 10-cm DBH and more than 1 m tall (snags) were inventoried on the entire 12-ha set of plots by measuring the basal and top diameters and height as well as assigning them to decay classes. Volume was determined for each species and decay class of logs and snags, and these were converted to mass by multiplying by species and decay class specific density values (Harmon and Sexton 1996). To determine variation at the scale of 1.0 ha, we computed the mean mass of logs and snags separately for each of the 12 ha that was sampled. Decomposition rate constants of logs were based on resampling 50 logs in 1998 originally sampled by Sollins (1982). The location of the logs was in the T. T. Munger RNA (Remillard 1999) and randomly within the crane plot. Samples were weighed in the field, and subsamples were dried at 55°C to determine moisture content. Stores in several other fine woody-detritus pools were estimated using assumptions based on the ratios of live parts, the relative decomposition rate of the parts, and their retention on dead boles. Dead coarse roots were estimated assuming they equaled 18%–26% of snag and log mass. This range was calculated by assuming that belowground woody tissues were the equivalent of 15%–20% of the aboveground woody biomass and then simulating the decomposition of the boles and roots at rates indicated by the field data for a 100-year period. The ratio for dead trees was then computed as the ratio of dead coarse roots and dead boles for this entire period. Suspended fine woody debris on snags was estimated using a similar set of calculations. In this case, dead attached branches were estimated to equal 10%–13% of the snag mass. As branches fall off of snags, we assumed that they were only attached to decay class 1 and 2 snags.
The decomposition of fine woody debris on the forest floor was measured by placing fresh branches of Douglas-fir and western hemlock on the forest floor and retrieving four branches of each species after 1, 2, and 3 years. Initial dry mass was determined by weighing the fresh branches and taking subsamples. Final dry mass was determined by removing the entire branches and oven drying them at 55°C. The decomposition rate constant of attached dead branches was estimated from the decomposition of four fresh branches each of Douglas-fir and western hemlock that had been suspended 1–2 m off the forest floor for 2 years in a similar experiment. The decomposition rate constant of dead coarse woody roots was estimated by excavating roots attached to stumps from trees that had been cut 4–50 years prior to excavation (Jansch 2001). A total of 21 Douglas-fir and 21 western hemlock stumps were sampled, with 8–24 roots excavated from each stump. The diameter and length of each root were measured to determine volume, and the dry mass was then determined after drying at 55°C (Chen and others 2001). The decomposition rate constant of each species was calculated using linear regression with a natural logarithmic transformation of root density against time since the tree was cut. The standard error used in uncertainty calculations was that of the regression model.

**Forest Floor.** The store of C in the forest floor [that is, excluding highly decomposed, buried coarse woody debris (CWD), but including partially and highly decomposed leaves, cones, and wood less than 1 cm in diameter] was determined by two methods. The first used a 5-cm-diameter, stainless-steel corer that was driven into the soil. The core was then extracted, and decomposed wood was separated from the other material. A total of 105 cores grouped in sets of five were taken throughout the T. T. Munger RNA systematically along the 21 transects used for understory biomass sampling. The second method sampled forest floor at the locations of the 10 soil pits by using five similar-sized cores. The forest-floor cores were taken above the sampling face of the soil pit and were pooled for each soil pit. In both cases, the samples were oven dried at 55°C and the ash content determined on a subsample by using a muffle furnace. The decomposition rate constant of the forest floor derived from fine litter and fine roots was estimated from 6-year-long litter decomposition experiments conducted at the H. J. Andrews Experimental Forest. Samples were removed after 1, 2, 4, and 6 years of decomposition. Litterbags were 20 × 20 cm and filled with 10 g of litter. At each sample time, four litterbags of each four species [Douglas-fir, western white pine (Pinus monticola), Pacific rhododendron (Rhododendron macrophyllum), and vine maple] were harvested and dried to constant mass at 55°C. To estimate the average rate of litter decomposition, we used long-term data to give year-specific decomposition rate constants. We extrapolated the decomposition rate between years 4 and 6 to estimate the long-term pattern of mass loss. We then simulated the accumulation of forest floor that would be expected if the inputs of fine litter were constant for a period of 50 years, and the ratio of input and this simulated store was used as an estimate of the average decomposition rate constant of the forest floor, excluding highly decomposed wood. We used the range of the four species to estimate the range of this parameter.

We also measured the amount of decomposed, brown-rotted wood buried within the forest floor. The methods used were as described above for the nonwoody material. Brownish red and highly decomposed wood was separated from the other types of forest-floor material. The decomposition rate constant of this extensively decomposed wood was not measured directly but assumed to encompass the range in decomposition rate constants observed for class 5 logs (that is, the most decayed).

**Dead Fine Roots.** We measured dead fine roots at the site by using the methods described for live fine roots; however, the recovery of dead fine roots less than 2 mm in diameter was very low (0.2 Mg C ha⁻¹ y⁻¹). This may have been caused by a misclassification of dead roots as live or fragmentation during the washing process. Regardless, if this was the correct mass of dead roots, the decomposition rate would have to be 20-fold higher than observed in temperate forests (Chen and others 2002). Although root bags probably underestimate decomposition rates, it is unlikely they do so by this amount. We therefore used the decomposition rate constant for dead fine roots and the mortality of fine roots to estimate the store in dead fine roots. The average decomposition rate constant for dead fine roots was estimated using results from a 4-year-long study from the H. J. Andrews Experimental Forest (Chen 1999). We used a procedure similar to that used for litter, extrapolating the decomposition rate constant for year 4 of the study. The results of four species [Douglas-fir, western hemlock, ponderosa pine (Pinus ponderosa), and red alder (Alnus rubra)] were used to give a range of possible values. It is highly likely that the rate constants for the Wind River will be similar to that found at the H. J. Andrews Experimental Forest, given that Chen and colleagues (2002) found very
little site-level variation within the forests of the Pacific Northwest.

**Mineral Soil.** The estimates of C stores in mineral soil are from Remillard (1999). Soil texture, the fraction of particles larger than 2 mm in diameter, bulk density, and C content were determined in 10 soil pits that were at least 1 m deep. The latter three variables were determined for three depths: (a) 0–20 cm, (b) 20–40 cm, and (c) 40–100 cm. The fraction of particles larger than 2 mm in diameter was estimated for each sample depth. Bulk density was determined from three 5-cm-diameter cores per depth. C content was determined from a well-mixed bulk sample of approximately 1000 mL taken from the entire depth of each sample zone. Although the lower depth of 100 cm is arbitrary, it represents the zone where most roots occur at this site; sampling to bedrock would have increased the total mineral soil store less than 25% (based on the observed decrease in C content with depth). Samples were air dried and sieved to remove organic as well as mineral particles larger than 2 mm in diameter. Aggregates larger than 2 mm were crushed and also analyzed. C content was determined by using a Leco 2000 C/N/S analyzer (Leco, St. Joseph, MI, USA). Soil C was then calculated based on the C content of all fractions, the bulk density, fraction of coarse particles, and depth. The integrated value to 100-cm depth for each of the soil pits was used to calculate the mean and standard error of soil C stores. We estimated the decomposition rate constant of mineral soil C by two methods. First, an upper bound was set by dividing mineral soil respiration by the mineral soil stores of C. Mineral soil respiration rates were calculated as the difference between total soil respiration and the sum of respiration associated with decomposing dead fine and coarse roots as well as that of the litter and rotten wood in the forest floor. Total soil respiration was determined from the mean efflux of soil CO$_2$ at eight locations near the canopy crane where a PVC (polyvinyl chloride)-pipe (5 cm deep and 10 cm in diameter) was installed 2 cm deep into the forest floor. CO$_2$-flux measurements were made in April, June, August, and October 1997 and January 1998 using a Li-Cor 6250 infrared gas analyzer (Li-Cor Biosciences, Lincoln, NE, USA) equipped with a soil respirometer. Second, a lower bound was set to 0.0025 y$^{-1}$, based on the results of a long-term laboratory incubation experiment (for example, see Hart and others (1994)) that were adjusted for temperature differences using a $Q_{10}$ of 2.

**Heart Rot.** In addition to the traditional components of heterotrophic respiration, we included an estimate of losses from heart rot within living trees. This respiration term was important to include, given 16.5% of the Douglas-fir trees at the site had fruiting bodies on their upper boles and many trees had dead tops, swellings at branch nodes, and other indications of heart rot and butt rot. Respiration associated with these rots was not measured directly. We used literature values from forests in the region to set an upper limit (25%) of stem wood volume being attacked by heart rot (Harmon and others 1996). The mass of heart rot was calculated as this fraction of bole wood, adjusted for past decomposition losses (heart rot was assumed to have half the density of sound wood). The lower limit was set at zero even though there is evidence to counter this assumption. We assumed that the rate constant of heart-rot decomposition was equal to the mean of logs.

**Carbon Content**

The C content of all pools except mineral soil was assumed to be 50%. For mineral soil, the values from the Leco 2000 C/N/S analyzer were used. For the forest floor, the C content of the non-ash portion of samples was assumed to be 50%. The majority of other tissues were woody, and a 50% C content is consistent with values found by Sollins and colleagues (1987) for sound and decayed wood.

**Heterotrophic Respiration ($R_h$)**

Heterotrophic respiration for each pool was calculated as the product of the store of detritus or mineral soil ($D$) and the decomposition rate constant ($k$):

$$R_h = kD$$

**Net Ecosystem Production (NEP)**

The net C flux at the site or NEP was calculated as the difference between NPP and $R_h$:

$$NEP = NPP - R_h$$

Because our method was based on multiple flux terms (some of which involve changes in stores), it should be considered an aggregated-flux approach. This was necessary because we did not have multiple measurements of the various C pools (aside from the live trees) required for the delta-stores approach.

**Uncertainty Analysis**

Rather than present one estimate of rates, we determined the uncertainty associated with these estimates. Given that our estimates represent long-term average rates and stores, we focused on uncertainty associated with spatial variation at the
approximate spatial scale measured by eddy-flux methods. Hence, our estimates of pool sizes and rate constants included an estimate of spatial variation at the level of multiple hectares, if possible. We also included uncertainty associated with measurements or estimates of pools or rate constants.

A Monte Carlo method was used to calculate the uncertainty of C stores, GPP, NPP, $R_a$, $R_b$, and NEP estimates. Here, the values of each flux, store, and each associated rate constant involved in calculations were varied from the mean ±2 SE. We assumed a normal distribution (that is, standard errors of the mean have a normal distribution). The standard error of the mean was used because it reflected the variation in the estimate of the mean at the scale of a hectare. The standard deviation, in contrast, would have represented the variation of individual samples, many with spatial extents far smaller than 1 ha. If the pools or the fluxes were estimated independently, then the value used for a pool or flux was selected independently for each round of calculations. If sizes or rate constants of one pool were used to derive the values of other pools, then variation in the former pool was directly linked to that of the latter. In the case of NEP, uncertainty calculations were done several ways. First, we assumed variation in NPP was independent of $R_b$. Second, we assumed that part of the variation in $R_b$ was associated with variation in NPP. This is because a major portion (more than 90%) of NPP for this stand was in the form of litterfall and mortality. When these fluxes vary, the store of detritus or soil C also varies and this influences $R_b$. We therefore linked a portion of the variation in detritus stores (equal to the fraction of NPP that was allocated to litterfall and mortality) to the variation in NPP. The remaining variation in detritus and soil stores varied randomly. Replicate Monte Carlo calculations were conducted, and the mean and standard deviation were computed using an Excel spreadsheet. The standard deviation was used in the final presentation of uncertainty, as it reflected the variation more liberally than did the standard error (which would have been 14–20 times lower, given the sample sizes used). In addition, as the Monte Carlo estimates were for the mean at the scale of a hectare, the standard deviation of these estimates is equivalent to the standard error at this scale (one can think of the standard error as the standard deviation of estimates of the mean). Uncertainty of each estimate was expressed as ±2 SD of 200–400 calculations, depending on the number required for the variance to stabilize.

**Results**

**Biomass**

The total store of C in living plants at the Wind River site was 39,800 g C m$^{-2}$ (34,800–44,800 g C m$^{-2}$) (Table 1). Trees form the majority of this store, comprising 99.2% of the total. Within trees, heartwood is the largest pool, and it comprises 39% of the site store in live plants.

**Net Primary Production**

The total change in live plant stores (that is, Δ stores) was 59 g C m$^{-2}$ y$^{-1}$ (11–107 g C m$^{-2}$ y$^{-1}$). Therefore, live stores might have increased over the period of observation as long as heart-rot losses were zero. Given that heart rot is present in the stand, this increase in live stores is likely to be overestimated by up to 46 g C m$^{-2}$ y$^{-1}$. The differences in change in C stores of live pools is similar to the distribution of live C stores. This observation must be tempered in that we assumed that fine-root, herb, shrub, and epiphyte C was not accumulating (that is, Δ stores was zero). If these other pools increased by the amount observed for trees (0.2% y$^{-1}$), then live stores would have increased an additional 0.5 g C m$^{-2}$ y$^{-1}$. Viewed another way, a 10% underestimate in the total change in Δ stores would occur only if nontree pools increased at a rate 10 times that of the tree pools (that is, 2% y$^{-1}$).

Mortality of plant parts was a major flux (Table 1), and at 523 g C m$^{-2}$ y$^{-1}$ (385–661 g C m$^{-2}$ y$^{-1}$) was approximately 10-fold larger than Δ stores and 25–50 times larger than grazing. The largest source of mortality was leaf fall, with 135, 26, and 40 g C m$^{-2}$ y$^{-1}$ for tree, shrubs, and herbs, respectively. The contribution from live branches, primarily in the form of twigs and cones, was 100 g C m$^{-2}$ y$^{-1}$. This high contribution of fine woody litter (32%) to total aboveground fine litterfall is quite typical of coniferous forest ecosystems in this region (Grier 1976; Grier and Logan 1977; Grier and others 1981). Fine-root mortality was 91 g C m$^{-2}$ y$^{-1}$, based on the observation that 25% of the fine roots were dying annually.

Grazing of tree foliage was low in 1995–96, amounting to 1%–8% of the current year’s foliage. Assuming an average damage value of 5%, and that all of this damage was due to herbivory, gives a grazing loss of approximately 10 g C m$^{-2}$ y$^{-1}$. Data from the visual survey of foliage damage at the site in 1999 indicated that 2% of the conifer foliage and 10% of the understory Acer circinatum leaves had damage. Assuming this fraction of all leaves was
eaten indicates the aboveground grazing flux was approximately 20 g C m\(^{-2}\) y\(^{-1}\). This range of values, equivalent of between 2% and 4% of total NPP, is typical of past estimates for conifer forests (Schowalter 1989).

Total NPP, including grazing, was estimated to be 597 g C m\(^{-2}\) y\(^{-1}\) (453–741 g C m\(^{-2}\) y\(^{-1}\)). Unlike \(\Delta\) stores, NPP was not related to the biomass distribution, with approximately 4% of the live C pools accounting for approximately 50% of the total NPP.

**Autotrophic Respiration and Gross Primary Production**

Annual \(R_a\) was estimated to be 1309 g C m\(^{-2}\) y\(^{-1}\) (845–1773 g C m\(^{-2}\) y\(^{-1}\)) (Table 2). This estimate does not include estimates for foliage “dark” respiration during the day. Aboveground parts accounted for 868 Mg g C m\(^{-2}\) y\(^{-1}\) or 66% of the total, and coarse and fine roots accounted for the other 441 g C m\(^{-2}\) y\(^{-1}\). The net assimilation (A) of foliage of conifers would have to have been 1123 Mg g C m\(^{-2}\) y\(^{-1}\) if the \(R_a\) associated with all tree components except leaves is added to tree NPP terms. Based on NPP and \(R_a\), GPP for this stand would have been 1906 g C m\(^{-2}\) y\(^{-1}\) (1444–2368 g C m\(^{-2}\) y\(^{-1}\)). This indicates that NPP is equivalent to approximately 31% of GPP.

**Detritus and Soil Pool Stores and Decomposition**

The total store of C in detritus and mineral soil was 22,100 g C m\(^{-2}\) (20,600–23,600 g C m\(^{-2}\)) (Table 3). Adding this store to the live C stores indicates that the Wind River old-growth forest stores a total of 61,899 g C m\(^{-2}\) (56,600–67,700 g C m\(^{-2}\)), with 36% of that comprised of detritus and mineral soil stores. The two largest pools were mineral soil and woody detritus, which stored 9300 and 9550 g C m\(^{-2}\), respectively. Fine, nonwoody litter was the smallest pool, storing 3180 g C m\(^{-2}\) of the total “dead” stores.

Decomposition rate constants for the detritus and mineral soil pools were highly variable, ranging from 0.005 y\(^{-1}\) for mineral soil to 0.191 y\(^{-1}\) for fine roots. Dividing the estimate of total \(R_h\) by the total soil and detritus store indicates an average decomposition rate constant of 0.026 y\(^{-1}\), which would correspond to a turnover time of 38 years.

**Heterotrophic Respiration**

Total \(R_h\), including respiration of grazers and heart rot, was estimated to be 577 g C m\(^{-2}\) y\(^{-1}\) (479–675 g C m\(^{-2}\) y\(^{-1}\)). The majority of the \(R_h\) flux was associated with the forest-floor litter, which, with a value of 187 g C m\(^{-2}\) y\(^{-1}\), accounted for 32% of the total flux with 8% of the C store. The next largest \(R_h\) flux was from woody detritus of all forms, with a value of 215 g C m\(^{-2}\) y\(^{-1}\) accounting for 37% of the total flux and slightly less than their proportion of C store (48% of total). Respiration of grazers, though not measured directly, was assumed to be equal to consumption, meaning 10–20 g C m\(^{-2}\) y\(^{-1}\) is respired by grazers. In our calculations of \(R_h\), we accounted for a heart-rot flux ranging from 0 to 46 g C m\(^{-2}\) y\(^{-1}\). This is small relative to the total estimated flux of \(R_h\), but is equivalent to 88%–176% of the estimated heartwood \(\Delta\) stores and indicates heartwood may not be accumulating in the stand as indicated by our NPP calculations.

**Net Ecosystem Production**

Using the NPP flux and \(R_h\) not corrected for heart-rot respiration indicates that the old-growth forest at Wind River is a C sink of +43 g C m\(^{-2}\) y\(^{-1}\) (since NEP is referenced to the ecosystem, a positive value indicates a sink and negative value indicates a source). Deducting the estimated range of losses from heart rot would make the stand either a slight source of –3 g C m\(^{-2}\) y\(^{-1}\) or a slight sink of +20 g C m\(^{-2}\) y\(^{-1}\). These results indicate that the Wind River old-growth stand is a slight sink for atmospheric C if heart rot in the stand is not too extensive. The maximum possible range in NEP including heart-rot losses, estimated by adding the lowest possible value of NPP to the highest possible of \(R_h\), is quite wide, bracketing a very moderate source to a very large sink (–222 to +262 g C m\(^{-2}\) y\(^{-1}\)). Monte Carlo-based estimates including heart-rot losses and assuming independent variation in NPP and \(R_h\) are more constrained, indicating –170 to +216 g C m\(^{-2}\) y\(^{-1}\) would be the most likely range. Partial dependence of NPP and \(R_h\) (reflecting the fact that 90% of NPP is in the form of mortality and litterfall) gives an even narrower range of –116 to +156 g C m\(^{-2}\) y\(^{-1}\). The latter range is the most likely, given the known dependence of detritus and soil stores on NPP and observation that most NPP at this site offsets losses to mortality.

**DISCUSSION**

**Carbon Stores**

Our estimate of total live biomass (39,807 g C m\(^{-2}\)) was slightly lower than the 43,500 g C m\(^{-2}\) found by Grier and Logan (1977) and considerably lower
than the approximately 73,500 g C m\(^{-2}\) reported by Means and colleagues (1992) for *Pseudotsuga*-dominated forests. Within the Pacific Northwest region, Smithwick and colleagues (2002) reported a range between 14,700 g C m\(^{-2}\) for east-side *Pinus*-dominated forests and 60,600 g C m\(^{-2}\) for coastal *Picea sitchensis*-Tsuga heterophylla forests, with a regional mean of 45,500 g C m\(^{-2}\). Boone and colleagues (1988) reported a live store of approximately 20,000 g C m\(^{-2}\) for a mature *Tsuga mertensiana* stand at high elevation, whereas Krumlik and Kimmins (1976) reported approximately 29,000 g C m\(^{-2}\) for an old-growth *Abies amabilis*-Tsuga mertensiana stand in British Columbia. Thus, our estimates lie within the range expected in this region.

Our estimates of detritus and mineral soil C stores (22,092 g C m\(^{-2}\)) also lie well within the range reported for the Pacific Northwest region. In *Pseudotsuga*-dominated forests, Grier and Logan (1977) and Means and colleagues (1992) reported 19,000 and 39,600 g C m\(^{-2}\), respectively, in detritus and soils. The range reported for the entire Pacific Northwest region by Smithwick and colleagues (2002) was from 7500 g C m\(^{-2}\) for east-side *Pinus*-dominated stands to 50,000 g C m\(^{-2}\) for coastal *Picea-Tsuga*-dominated forests. The regional mean detritus and mineral soil store from this study is 27,500 g C m\(^{-2}\). High-elevation *Tsuga mertensiana* forests were estimated to store 10,000 g C m\(^{-2}\) in detritus and mineral soil (Boone and others 1988). The proportion of total C stored in detritus and mineral soils at Wind River is close to the regional mean of 38%; however, there is considerable range in this proportion, with the highest proportions at coastal sites (45%) and high elevation (47%) (Boone and others 1988; Smithwick and others 2002).

Our estimates of total C stores (61,899 g C m\(^{-2}\)) also lie within the range reported in the Pacific Northwest. Grier and Logan (1977) and Means and colleagues (1992) report a total C store of 62,400 and 111,000 g C m\(^{-2}\), respectively, with the value from Means and colleagues being just below the absolute upper limit of 122,400 g C m\(^{-2}\) found by Smithwick and coworkers (2002). The regional mean from Smithwick and colleagues was 73,300 g C m\(^{-2}\), with the regional maximum of 110,700 g C m\(^{-2}\) found in coastal *Picea-Tsuga* forests and the regional minimum of 22,200 g C m\(^{-2}\) found in *Pinus*-dominated east-side forest. Total C stores at Wind River are considerably higher than the 21,500 g C m\(^{-2}\) found in high-elevation *Tsuga mertensiana* forests (Boone and others 1988).

Net Primary Production, Autotrophic Respiration, and Gross Primary Production

There are few ecosystem estimates of total NPP in the Pacific Northwest region to compare with our estimates; however, values for the most similar ecosystem are 544 g C m\(^{-2}\) y\(^{-1}\) (Grier and Logan 1977) and within our estimated range (453–741 g C m\(^{-2}\) y\(^{-1}\)). Both estimates are well below the 891 g C m\(^{-2}\) y\(^{-1}\) reported by Gower and colleagues (1992) for a 50-year-old *Pseudotsuga* forest in New Mexico. This difference might be caused by differences in forest age, as NPP is known to decrease once forest canopies have closed (Ryan and others 1997; Acker and others 2000, 2002), or alternatively it may be caused by different methods of estimating the belowground NPP. For example, the root mortality rates reported by Gower and coworkers (1992) indicate that roots have a mean life time of 0.75 years, whereas our estimates indicate a mean lifetime of 3–5 years. Total NPP at Wind River was also lower than the 840 g C m\(^{-2}\) y\(^{-1}\) found at a 180-year-old *Abies amabilis* stand in Washington (Grier and others 1981) or the 650 g C m\(^{-2}\) y\(^{-1}\) for aboveground NPP of coastal *Picea-Tsuga* forests (Grier 1976). Although the latter is an underestimate of total NPP, the difference in aboveground NPP with the Wind River site (444 g C m\(^{-2}\) y\(^{-1}\)) is consistent with the 1.5-fold greater store in live biomass at the coastal site. The lower value of total NPP than at the high-elevation *Abies*-dominated site is most likely due to the lower fine-root NPP estimates at Wind River, as indicated by the mean lifetime of fine roots of 1.1 years at the *Abies* stand. Aboveground NPP at the high-elevation *Abies*-dominated site was 228 g C m\(^{-2}\) y\(^{-1}\) and is more in line with the difference in live C stores found between the sites.

Our estimate of $R_a$ for Wind River of 1309 g C m\(^{-2}\) y\(^{-1}\) is very preliminary. It is far lower than the 7500 g C m\(^{-2}\) y\(^{-1}\) reported by Grier and Logan (1977) and roughly twice the 765 g C m\(^{-2}\) y\(^{-1}\) predicted by Kaduk and colleagues using a model. Measurements of live pools other than leaves at the site would help resolve the latter difference. Because of the vast differences in $R_a$ between our study and that of Grier and Logan (1977), our estimate of GPP is 7.7-fold smaller than theirs. Our estimate of GPP is lower than the annual value of 2200–2460 g C m\(^{-2}\) y\(^{-1}\) estimated by Winner and colleagues (2004) using the Soil-Plant-Atmosphere model, but larger than the 1300 g C m\(^{-2}\) y\(^{-1}\) modeled by Kaduk and colleagues (2004) and the 1570 g C m\(^{-2}\) y\(^{-1}\) estimated by Paw U and colleagues (2004) from eddy flux.
Net Ecosystem Production

The only other published ground-based estimates of old-growth *Pseudotsuga* forest NEP are by Grier and Logan (1977), who reported a C sink of 166 g C m$^{-2}$ y$^{-1}$. However, this estimate must be interpreted in light of uncertainties in $R_h$ (identified by Grier and Logan) that have been resolved (Franklin and others 1987; Harmon and Chen 1991; Chen and others 2001; Harmon and others 2001). The most significant improvement has been information on the rate that woody detritus decomposes. Grier and Logan assumed a decomposition rate constant for this material of 0.0067 y$^{-1}$, based on the ages of trees found growing on several logs. Since that time, higher rate constants, similar to the ones we used, have been measured by a variety of studies (Harmon and others 2001). This indicates that the Grier and Logan (1977) estimate of $R_h$ needs to be increased by 120 g C m$^{-2}$ y$^{-1}$, and thus their overall estimate of NEP would be 44 g C m$^{-2}$ y$^{-1}$. Grier and Logan also assumed, given the lack of root decomposition data at the time, that $R_h$ associated with dead roots would equal 50% of root mortality. Subsequent work on fine-root and coarse-root decomposition has found no basis for this assumption (Chen and others 2001, 2002). Other factors might also decrease the Grier and Logan (1977) NEP estimate. For example, they noted that heart rot was present in the stand but did not include these respiration losses. As little as 10% of heart rot in stems of their forest could completely offset the gains in stem stores they estimated. These values of heart rot are within the range typical for old-growth *Pseudotsuga* forests (Harmon and others 1996).

Unless some form of long-term production or removal from the forest has been neglected by our measurements, the uncertainty analysis of long-term ground-based measurements indicates that it is unlikely that the stand at Wind River is as strong a long-term sink as estimated by the eddy-covariance method in 1998–99 [150–220 g C m$^{-2}$ y$^{-1}$ (Paw U and others 2004)]. The error in ground-based measurements would have to be at least 100 and possibly as large as 200 g C m$^{-2}$ y$^{-1}$. Our uncertainty analysis indicates that approximately 95% of the long-term ground-based estimated values lie below +150 g C m$^{-2}$ y$^{-1}$.

Assuming that the eddy-flux tower system and related calculations are not resulting in underestimates in respiration terms, there are several hypotheses that might explain the differences between our ground-based and eddy-covariance-based estimates, including (a) undetected biomass increases, (b) underestimates of NPP, and (c) unmeasured C losses from the system in the ground-based system, as well as (d) a temporal mismatch between the long-term ground-based versus short-term flux tower measurements. Each hypothesis is addressed below.

The first hypothesis is that biomass increases were not detected by our measurements. However, measurements of diameters and heights in nearby permanent plots by using individually tagged trees indicate that live C stores have steadily increased by 920 g C m$^{-2}$ over the last 50 years, giving an annual increase of 18 g C m$^{-2}$ y$^{-1}$ (Bible 2001). These increases do not account for losses from heart rot, however, and therefore are likely to be overestimated. Although our estimates include root biomass, it is possible that more C is being allocated to coarse roots than in the past. If the relationship between aboveground parts and coarse roots has changed since the 1970s, this difference should be evident from a comparison of root to shoot ratios of trees sampled in the 1970s and today. Unfortunately, data for such a comparison do not currently exist. Another possibility is an increase in CWD stores; however, a comparison with the data collected 20 years earlier at the Wind River old-growth stand by Sollins (1982) does not support this hypothesis. Sollins estimated a mean C store of 6750 g C m$^{-2}$, whereas our mean estimate was 7080 g C m$^{-2}$. Although these measurements were not taken on the same plots, they were in very close proximity and differed by less than 0.5 SE. Using current estimates of decomposition rate constants and long-term mortality records from plots near the stand we examined (Bible 2001) indicates that even if CWD C stores were zero 50 years ago, CWD could only account for a sink of 50 g C m$^{-2}$ y$^{-1}$. A more reasonable initial CWD store of 75% of the current value yields a current potential sink of 16 g C m$^{-2}$ y$^{-1}$. C stores in mineral soil could also be increasing, but difficult to detect. By only sampling to 1 m depth, we probably underestimated the mineral soil C store by 25%, based on decreases in C content with soil depth. This would underestimate the total C store of the site by 3.7%, but assuming these deeper layers respired at the same rate constant as the shallower soil would also underestimate $R_h$ by 2% and, hence, increase the difference between the eddy-flux and the ground-based estimates. Our estimates of the decomposition in the mineral soil C pool are highly uncertain; if they are high, this could lead to an increased NEP estimate. C increase in mineral soil with sandy texture during old-field succession was approximately 4 g C m$^{-2}$ y$^{-1}$ (Richter and others...
1999), although this rate should be higher on soils with more clay content, as found at Wind River (Hassink and Whitmore 1997). Globally, Schlesinger (1990) considered an increase of 20 g C m\(^{-2}\) y\(^{-1}\) to represent the upper limit of soil C increases. With the higher of these two rates, most of the current C store in mineral soil could have accumulated since the last major disturbance approximately 450 years ago. Given that disturbance is thought to have a minimal effects on C in mineral horizons of forests (Johnson and Curtis 2001), it seems unlikely that C accumulation in the mineral soils is causing a major underestimate in NEP.

The second hypothesis is that NPP was underestimated. Grazing losses are difficult to measure, but because grazing increases both NPP and \( R_h \) it should not influence the NEP estimate unless grazer C is increasing. Our estimates of grazing losses and respiration of consumers are admittedly crude, but within the range observed by others (Schowalter 1989). For grazing to account for a systematic underestimate of NEP large enough to explain the eddy-flux versus ground-based discrepancy, an amount equivalent to 20% of the foliage would have been grazed, and most of that C would have to be accumulating in grazers each year. The most likely grazers are insects and, because these are short-lived organisms, they store very little C, with a range of 190–210 g C m\(^{-2}\) in Pacific Northwest forests (Schowalter 1989). The required accumulation of insect C implies that insect C is increasing several fold annually. This increase is also unlikely for large, longer-lived vertebrates such as deer and elk. Another component of NPP that we did not measure was in DOC associated with throughfall and stem flow. Grier and Logan (1977) estimated this flux to be 30 g C m\(^{-2}\) y\(^{-1}\), and a similar value would be expected at Wind River. These are likely to be highly decomposable materials, however, and the majority should be respired within a year of deposition. This flux is likely to increase both NPP and \( R_h \) and have little net effect on the NEP estimate. Losses via biogenic hydrocarbon emissions could also lead to underestimates of NPP, although given the small size of this flux [that is, 0.5%–2% of CO\(_2\) fluxes (Winner and others 2004)] it would also have little effect on our NEP estimate.

The third hypothesis is that we failed to measure a C export from the ecosystem. As we did not measure DOC losses from the stand at Wind River, this is one logical source of bias. These losses have been measured at the H. J. Andrews Experimental Forest as 3 g C m\(^{-2}\) y\(^{-1}\) (Grier and Logan 1977; Swanson and others 1982). Typical concentrations of DOC extracted from mineral soil in the region by tension and tension-free lysimeters are 1–10 Mg C L\(^{-1}\) (K. Lajtha personal communication). Assuming Wind River old-growth evapotranspires the same as basins in the H. J. Andrews Experimental Forest means that \( 1.6 \times 10^4 \) m\(^3\) ha\(^{-1}\) of water leaves the Wind River stand via groundwater, and using the mineral soil DOC values observed by Lajtha yields an annual export of DOC of 1–10 g C m\(^{-2}\) y\(^{-1}\), similar to the magnitude reported by Grier and Logan (1977). It is also possible that particulate organic C (POC) is leaving via erosion and fluvial transport. Estimates of POC exports via litterfall and an assortment of geomorphic processes in steep slopes in Pacific Northwest are approximately 2 g C m\(^{-2}\) y\(^{-1}\) (Swanson and others 1982). It is likely that on the flat terrain at Wind River rates of POC export would be lower, indicating omitting this flux would cause minimal underestimates in NEP. Transport of larger material such as logs in the seasonal stream running through the crane plot is also highly unlikely and has not been observed since the plot was established.

The final hypothesis involves the fact that the flux tower measured a single year, whereas we calculated a long-term average. For the most part, our estimates of NPP and \( R_h \) are based on long-term measurements with no estimate of annual variation. There are, however, two exceptions. Wind River site litterfall has a year-to-year variation of \( \pm 30\% \). Litter decomposition rate constants between the driest and wettest year at H. J. Andrews Experimental Forest vary by \( \pm 35\% \) (M. Harmon unpublished data). Assuming that all NPP components vary the same as litterfall, NPP is likely to vary within a range of 418–776 g C m\(^{-2}\) y\(^{-1}\). Similarly, by assuming that \( R_h \) varies as much as short-term litter decomposition, this flux is likely to vary from 404 to 750 g C m\(^{-2}\) y\(^{-1}\). Using Monte Carlo methods to vary these two fluxes and assuming these two fluxes vary independently of each other yields an approximate NEP range of \( \pm 227 \) to \( \pm 267 \) g C m\(^{-2}\) y\(^{-1}\). Thus, annual variation is the hypothesis most consistent with the value estimated by the eddy-flux method and therefore warrants additional examination.

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